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# Effects of nitrogen fertilizer, soil temperature and moisture on the soilsurface $CO_2$ efflux and production in an oasis cotton field in arid northwestern China

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## ABSTRACT

In several studies of agricultural ecosystems, researchers have focused on the soil-surface carbon dioxide  $(CO_2)$ effluxes, but the nature of CO2 production in the soil profile and its influencing factors remain unclear. In this study, the soil-surface CO<sub>2</sub> effluxes in an oasis cotton field were measured using the chamber method, and the CO<sub>2</sub> concentrations were used to estimate the CO<sub>2</sub> production in different layers of the soil profile using the gradient method. The soil CO2 concentrations increased with increasing soil depth, whereas CO2 production decreased with increasing soil depth. Both soil-surface CO2 effluxes and CO2 production in the 0-40 cm layers exponentially increased with increasing temperature. Irrigation temporarily reduced the soil-surface CO2 effluxes by 19–63% through inhibiting CO<sub>2</sub> production in the 10–40 cm layer but did not affect the CO<sub>2</sub> production in the 0-10 cm layer. CO<sub>2</sub> production mainly occurred in the 0-10 cm layer, and this cumulative production accounted for 63-67% of the total production throughout the soil profile (0-40 cm). The application of nitrogen (N) fertilizer enhanced the rate of  $CO_2$  production in the 0–20 cm layer by increasing the root biomass and soil mineral N content. A positive correlation was detected between the soil-surface CO<sub>2</sub> efflux and soil NO<sub>3</sub><sup>-</sup> content in 2015, but no significant correlations were found between the soil-surface CO<sub>2</sub> efflux and soil NH<sub>4</sub><sup>+</sup> contents in any treatment. A higher soil-surface CO<sub>2</sub> efflux was observed under high soil temperature and a certain soil moisture range (0.21–0.23 cm<sup>3</sup> cm<sup>-3</sup>). An analysis of the soil profile revealed higher CO<sub>2</sub> production rates detected in the 0–10 cm layer under high soil temperature and moisture conditions, but higher rates were observed under high soil temperature and low soil moisture conditions in the 10-20 cm layer. Therefore, our results suggest that the effects of fertilization, soil temperature and moisture on CO<sub>2</sub> production vary depending on the soil depth. These findings might improve our understanding of the mechanisms underlying soil respiration in soil profiles.

## 1. Introduction

In terrestrial ecosystems, soil is the largest carbon (C) pool, containing approximately 2400 Pg C in the upper 2 m (Batjes, 1996). Soilsurface carbon dioxide (CO<sub>2</sub>) efflux is one of the largest C fluxes (Schimel, 1995) that results in a net loss of C to the atmosphere, mainly through soil respiration and the combination of root and heterotrophic respiration (Hanson et al., 2000). This loss might increase with increasing temperatures through stimulation of biological activity in the soil (Gaumont-Guay et al., 2006), and the released CO<sub>2</sub> might form part of a positive feedback by contributing to climate warming. Soil temperature and moisture are the major abiotic factors controlling soil

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respiration through their effects on soil biological activities and the decomposition of soil organic matter (Gaumont-Guay et al., 2006; Koncz et al., 2015a). Soil respiration is also strongly linked to plant biomass by influencing the translocation of photosynthate during rhizosphere respiration (Ding et al., 2010; Koncz et al., 2015b; Scheer et al., 2013).

The soil-surface  $CO_2$  efflux is the combined result of  $CO_2$  production and transport between different soil layers (Rey, 2015), and many researchers have focused on this efflux using chamber method (Brumme and Beese, 1992; Lv et al., 2014; Mosier et al., 2006; Wang et al., 2013) and have assumed that it is equivalent to soil respiration. However, this measured soil-surface  $CO_2$  efflux is not likely to represent the "real" soil







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respiration and thus provides limited information on the CO<sub>2</sub> dynamics within the soil profile (Rey, 2015). For example, when the soil moisture exceeds the soil water field capacity, the soil-surface CO<sub>2</sub> efflux generally decreases with increasing soil water content (Gaumont-Guay et al., 2006). This result not only reflects a reduction in soil respiration but also results from a restriction of CO<sub>2</sub> transport in the soil profile. Moreover, due to the variety of root and soil organic matter, the effects of soil temperature and moisture on CO<sub>2</sub> production rates likely vary depending on the soil depth. Therefore, understanding the CO<sub>2</sub> production in the soil profile is important for improving our understanding of the interactions between influencing factors and "real" soil respiration. Several researchers have measured the soil CO<sub>2</sub> concentrations at different depths to determine the production of CO<sub>2</sub> in the soil profile using the gradient method (Nan et al., 2016; Pumpanen et al., 2008; Vargas and Allen, 2008), which can be used to investigate the "real" soil respiration in the soil profile.

In agricultural ecosystems, the application of nitrogen (N) fertilizer not only supplies nutrients to improve crop growth but also affects soil respiration (Bhattacharyya et al., 2012; Ding et al., 2010; Fuß et al., 2011). In general, N fertilization increases soil respiration due to increased root respiration resulting from an increase in root biomass (Bhattacharyya et al., 2012; Fuß et al., 2011; Shao et al., 2014) and also increases heterotrophic respiration by increasing the decomposition of soil organic C (SOC) by reducing the C/N ratio (Kuzyakov et al., 2000; Luo et al., 2016). However, in a maize field, Ding et al. (2010) found that the application of N fertilizer reduced soil respiration, because the N uptake needed for maize growth in a non-fertilized plot could be approximately met by decomposing SOC, which resulted in higher root respiration compared with that in a fertilized field. In a cotton field, Liu et al. (2008) reported that the root biomass increased with increasing N fertilization rates, which likely enhanced soil respiration, because root respiration was the main component of the total soil respiration (Yu and Zhao, 2015).

Over the past 20 years, the production of cotton in China has increased by 26% (Statistics of the Food and Agriculture Organization of the United Nations (FAOSTAT), 2015), and these increases are largely driven by the intensification of agricultural management (e.g., N fertilization). The arid region of China is an important area for cotton production, and the cotton lint produced in this region represents 60% of the total production of cotton lint in China (China Statistical Yearbook, 2014). In this region, soil-surface CO<sub>2</sub> effluxes in agricultural systems are two- to five-fold higher than those in natural ecosystems (Lai et al., 2012). Moreover, the fertilizer rates in arid cotton fields range from 240 to 360 kg N ha<sup>-1</sup> yr<sup>-1</sup> (Lv et al., 2014), making them higher than those in other regions, including Northern China, where fertilizer rates typically range from 60 to 80 kg N ha<sup>-1</sup> yr<sup>-1</sup> (Liu et al., 2014). A previous study showed that both root and heterotrophic respiration in a cotton field increased with increasing N fertilization based on the DeNitrification-DeComposition (DNDC) model (Yu and Zhao, 2015), but this result has not been validated by experimentation in the field. Therefore, with the heavy use of N fertilizers coupled with high soil respiration rates, it is necessary to understand the links between N inputs and soil respiration in this agricultural ecosystem. In this study, we used the chamber method to measure the soil-surface CO<sub>2</sub> efflux and the gradient method to estimate CO<sub>2</sub> production under different N fertilizer treatments. The objective was to assess the effects of soil temperature and moisture on the soil-surface CO<sub>2</sub> efflux and CO<sub>2</sub> production rates, and we hypothesized that the application of N fertilizer would increase both soil-surface CO<sub>2</sub> efflux and the production of  $CO_2$  in the soil profile.

#### 2. Materials and methods

## 2.1. Site and experiment description

The field experiment was conducted during two growing seasons

(2014 and 2015) at the Aksu National Experimental Station in the Oasis Farmland Ecosystem, which is located in north-western China (40°37′N, 80°45′E; elevation: 1028 m). This region has a typical arid climate, with an annual mean air temperature of 11.2 °C and annual mean precipitation of 45.7 mm. At the soil layers of 0–10, 10–20 and 20–40 cm, the soil organic matter contents are 8.0, 4.8 and 3.8 g C kg<sup>-1</sup>, respectively, and the C/N ratios are 11.9, 11.1 and 12.6. The soil is gleyic solonchak (World Reference Base for Soil Resources), and the soil texture is silt loam with 6% clay (< 0.002 mm), 43% silt (0.002–0.02 mm) and 51% sand (0.02–2 mm) in the 0–40 cm layer, with bulk densities of 1.49, 1.52 and 1.56 g cm<sup>-3</sup> at the 0–10, 10–20 and 20–40 cm soil depths, respectively.

Before sowing, basal fertilization and tillage occurred on 13 April 2014 and 15 April 2015. Basal fertilizers were incorporated into the 0-30 cm soil layer at rates of  $80 \text{ kg N} \text{ ha}^{-1} \text{ year}^{-1}$  in the form of urea (60 kg N ha<sup>-1</sup> year<sup>-1</sup>), diammonium phosphate (20 kg N ha<sup>-1</sup> year<sup>-1</sup>) and potash followed by tillage. After two days, ridge soils were formed (100 cm wide and 5 cm high) by a ridging plough, and plastic film (0.02 mm thick and 1.2 m wide) was mulched over the ridged soil. The edges of the film were sealed under the soil, and the furrow soil (50 cm wide) remained uncovered. Cotton (Tanong No. 8) seeds were sown into the mulched ridges at 10 cm intervals in rows spaced 50 cm apart on 15 April 2014 and 17 April 2015, and in both years, drip irrigation was used to supply 320 mm of irrigation water in 8 doses of 40 mm each. During the drip irrigation period, three fertilization treatments were applied: 0, 160 and 320 kg N ha<sup>-1</sup>. For the 160 and 320 kg N ha<sup>-1</sup> treatments, urea was uniformly dissolved in the irrigation water and applied as four doses of 40 and 80 kg N ha<sup>-1</sup>, respectively, to improve crop N uptake in this plastic mulched cropping system (Wang et al., 2016). Urea application occurred on 9 July, 20 July, 28 July and 6 August in 2014 and on 5 July, 13 July, 21 July and 28 July in 2015 (Fig. 1b). Therefore, the total N fertilizer rates were 80, 240 and 400 kg N ha<sup>-1</sup>, but these treatments are denoted 0, 160 and  $320 \text{ kg N} \text{ ha}^{-1}$ , respectively, during the observation period. Each treatment (10 m long and 6 m wide) was assigned in a completely randomized design and replicated three times.

## 2.2. Sampling

Three gas samples per plot at depths of 10, 20 and 40 cm were collected from the ridge soil profiles between cotton plants every four to ten days in 2014 (from 18 May to 11 November) and 2015 (from 8 May to 12 October) using modified diffusion equilibrium samplers. At the beginning of the experiment, 27 gas collectors were installed in the ridge soil. Each collector was made of a Teflon tube (1.0 mm inner diameter and 3.0 mm outer diameter) connected to a probe for sampling the soil gas. The probe consisted of a 60 ml PVC pipe (12 mm inner and 15 mm outer diameters) with 16 holes (2 mm inner diameter), and the bottom was sealed with a glass microfiber filter to allow soil air to diffuse into the sampler. At each sampling position, a soil auger (2 cm inner diameter and 15 cm in length) was used to excavate a hole in the centre of four cotton plants. The gas collectors were installed vertically into the soil to collect gas samples at depths of 10, 20 and 40 cm in each plot, and each collector was separated horizontally by approximately 20 cm. The equipment was left in the field for approximately half a month to allow the soil CO<sub>2</sub> concentration to reach equilibrium before sampling. To collect gas from each depth, we used a gastight, three-way ball valve fitted with an injection syringe, a design that enabled us to extract soil air at any depth without contamination or clogging. After pre-extraction of the residual air, a 30 ml gas sample was collected using a syringe. The air CO<sub>2</sub> concentrations were also measured from the ambient gas above the soil surface (0 cm).

The soil-surface  $CO_2$  efflux from each plot was measured in the ridge soil between cotton plants using the closed-chamber method and calculated using either the non-linear or linear methods described by Wang et al. (2013). The equipment consisted of a stainless-steel chamber (30 cm length, 15 cm width and 15 cm height) and a stainless-



**Fig. 1.** Soil temperature (a), moisture and amount of rainfall or irrigation (b), and  $CO_2$  diffusion coefficient (c) at 10, 20 and 40 cm depths in 2014 and 2015. Black arrows (b) denote the application of urea. *Error bars* for the different variables represent the standard errors of the means (n = 3).

steel base (30 cm length, 15 cm width and 5 cm height). Each chamber was covered with a foam plate and tinfoil to limit the temperature increase within the chamber during sampling. The base was vertically inserted 5 cm into the soil. During the gas sampling, the flange of each chamber was carefully placed in the groove of the base, and each groove was filled with an appropriate amount of water to ensure a closed environment. The gas samples were collected between 11:00 and 13:00. After chamber closure, five gas samples (60 ml) were sampled at 0, 10, 20, 30 and 40 min from the headspace with a syringe. The gas samples were then analysed using a modified gas chromatograph (Agilent 7890A, Agilent, Palo Alto, CA, USA) equipped with a nickel catalyst converter to reduce  $CO_2$  into  $CH_4$ , and the  $CO_2$  concentration was then measured using a hydrogen flame ionization detector (FID) at 375 °C. A standard gas with a known  $CO_2$  concentration (398.9 ppm  $CO_2$  in N<sub>2</sub>) was used to calibrate the concentration of the  $CO_2$  samples.

Six plants were sampled in each plot at the end of the growing season (on 4 October 2014 and 28 September 2015). The aboveground portions (leaves and stems) of the cotton plants were randomly collected using pruning shears, and roots were collected using a root auger (5 cm inner diameter and 20 cm length) from the 0–40 cm layer because most roots are distributed in this layer (Zhao et al., 2010). All roots were washed and separated by hand. The samples were ovendried at 80 °C for > 48 h and weighed. The soil temperatures at different layers (0–10, 10–20 and 20–40 cm) were measured directly using Hydra Probes (Hydra Probe II, Stevens Water Monitoring Systems Inc., Portland, OR, USA). Soil samples were collected from different layers (0–10, 10–20 and 20–40 cm) and oven-dried at 105 °C for > 24 h to determine the soil water content.

One soil sample in the 0–10 cm layer per plot was collected using a soil auger (2 cm inner diameter and 15 cm length), which was used to measured soil nitrate  $(NO_3^{-})$  and ammonium  $(NH_4^{+})$  contents. The soil

was sieved (2 mm mesh), and 5.0 g of soil was then extracted with 100 ml of 0.01 M  $CaCl_2$  solution on a mechanical shaker for 1 h. The extracts were analysed with an automated  $NO_3^-$  and  $NH_4^+$  analyser (AutoAnalyser 3, Bran Luebbe/SEAL Analytical, Norderstedt, Germany).

## 2.3. Calculation of the production of $CO_2$ in the soil profile

We calculated CO<sub>2</sub> production using the gradient method, which is based on the CO<sub>2</sub> concentrations and transport properties of the soil profile (Maier and Schack-Kirchner, 2014). The CO<sub>2</sub> production (g C m<sup>-2</sup> s<sup>-1</sup>) between the depths i + 1 and i cm ( $P_{i + 1,i}$ ) was calculated as follows:

$$P_{i+1i} = F_{ii-1} - F_{i+1i} \tag{1}$$

where  $F_{i,i-1}$  and  $F_{i+1,i}$  are the soil CO<sub>2</sub> effluxes (g C m<sup>-2</sup> s<sup>-1</sup>) from depths i - 1 to i cm and from depths i + 1 to i cm, respectively. Moreover, the soil-surface CO<sub>2</sub> efflux ( $F_0$ ) and the CO<sub>2</sub> efflux from depths 10 to 0 cm ( $F_{10_0}$ ) were used to calculated CO<sub>2</sub> production ( $P_{10_0}$ ) in the 0–10 cm layer:

$$P_{100} = F_0 - F_{100} \tag{2}$$

In the soil profile, we assumed that the  $CO_2$  transport in the soil was dominated by molecular diffusion and that the  $CO_2$  efflux from depths *i* + *1* to *i* could be calculated based on the effective gas diffusivity and the  $CO_2$  concentration gradient using Fick's law in one dimension:

$$F_{i+1,i}(z) = -D_s \frac{dC}{dz}$$
(3)

where  $F_{i+1,i}(z)$  is the CO<sub>2</sub> efflux (g C m<sup>-2</sup> s<sup>-1</sup>) from depths i + 1 to i;  $D_s$  is the effective soil gas diffusivity in the soil (m<sup>3</sup> soil air m<sup>-2</sup> soil s<sup>-1</sup>); C is the CO<sub>2</sub> concentration (g C m<sup>-3</sup>); z is

the depth (m); and dC/dZ is the soil CO<sub>2</sub> concentration gradient in the soil profile.  $D_s$  can be estimated from the structure-dependent, water-induced linear reduction model (Moldrup et al., 2013). Fan and Jones (2014) suggested that the following equation is a reasonable method for determining the gas diffusion coefficient used for estimating CO<sub>2</sub> efflux:

$$D_{s} = D_{0}\varepsilon^{(1+C_{m}\varnothing)}\left(\frac{\varepsilon}{\varnothing}\right)$$
(4)

where  $D_0$  is the gas diffusion coefficient in air (m<sup>2</sup> s<sup>-1</sup>);  $\varepsilon$  is the soil air content (m<sup>3</sup> m<sup>-3</sup> soil);  $\Phi$  is the soil porosity (m<sup>3</sup> m<sup>-3</sup>); and  $C_m$  is the media complexity factor. Moldrup et al. (2013) recommended a  $C_m$  value of 2.1 for intact soil, and  $\varepsilon$  and  $\Phi$  were computed as follows:

$$\emptyset = 1 - \frac{\rho_b}{\rho_s} = \varepsilon + \theta \tag{5}$$

where  $\rho_b$  is the dry bulk density (g m<sup>-3</sup>) of different soil layers;  $\rho_s$  is the soil particle density (2.65 g m<sup>-3</sup>); and  $\theta$  is the soil water content at different depths (m<sup>3</sup> m<sup>-3</sup>). The  $D_0$  of CO<sub>2</sub> in the atmosphere was calculated as follows:

$$D_0 = D_{stand} \left(\frac{T}{T_0}\right)^{1.75} \left(\frac{P_0}{P}\right)$$
(6)

where  $D_{stand}$  is a reference value  $(1.39 \times 10^{-5} \text{ m}^2 \text{ s}^{-1})$  at  $T_0$  (293.15 K);  $P_0$  (1013 hPa) is the CO<sub>2</sub> gas diffusion coefficient in atmospheric air (Pritchard and Currie, 1982); *T* is the air temperature (K); and *P* is the pressure at Aksu station (892 hPa). Finally, the units of time for soil CO<sub>2</sub> production were converted from per second (s<sup>-1</sup>) to per day (day<sup>-1</sup>).

## 2.4. Data analysis

All statistical analyses were conducted using the SPSS v. 16.0 software package (SPSS, Chicago, IL, USA). We used a repeated-measure analysis of variance (ANOVA) to test the differences in soil temperature, moisture, gas diffusion coefficient, daily soil-surface  $CO_2$  efflux and  $CO_2$  production among the three soil layers or three fertilizer treatments, and the differences were tested at a level of p < 0.05. The cumulative soil-surface  $CO_2$  efflux and  $CO_2$  production in the soil profile were estimated by linear interpolation of the measured daily values with the corresponding time period; the results were then summed over the observation period. One-way ANOVA was used to analyse the differences in the cumulative soil-surface  $CO_2$  efflux, cumulative  $CO_2$  production in the soil profile, root and total biomass of cotton among the three fertilizer treatments, and the differences were tested using least-significant differences (LSDs) at a level of p < 0.05.

A linear regression analysis was performed to fit the relationship between soil-surface CO<sub>2</sub> efflux and soil mineral N (NO<sub>3</sub><sup>-</sup> and NH<sub>4</sub><sup>+</sup>) content, and the Q<sub>10</sub> model by Gaumont-Guay et al. (2006) was used to describe the effects of soil temperature on soil-surface CO<sub>2</sub> efflux and  $CO_2$  production in the soil profile. When analysing the effect of soil temperature on soil-surface CO<sub>2</sub> efflux or CO<sub>2</sub> production, we excluded the data collected immediately after irrigation because excessive soil moisture (>  $0.30 \text{ cm}^3 \text{ cm}^{-3}$ ) depresses soil respiration (Gaumont-Guay et al., 2006). A hyperbolic regression was used to model the relationship between soil-surface CO<sub>2</sub> efflux and soil moisture (Gaumont-Guay et al., 2006), and a linear regression was used to fit the relationships between CO<sub>2</sub> production and soil moisture. Multiple regressions were used to fit the effects of soil temperature and moisture on soil-surface CO2 efflux or CO2 production. Lastly, we used the coefficient of determination (R<sup>2</sup>), the root mean square error (RMSE) (Gaumont-Guay et al., 2006) and Akaike's information criterion (AIC) values (Burnham and Anderson, 2004) to select the best regression model to describe the effects of soil environmental variables on soil-surface CO<sub>2</sub> efflux or CO<sub>2</sub> production in the soil profile. The model with higher R<sup>2</sup> values and lower RMSE and AIC values was considered to be strongly supported.

#### 3. Results

#### 3.1. Soil properties and crop biomass

During the observation period, the soil temperature at different depths remained high from May to August and then gradually decreased (Fig. 1a), and no significant differences in soil temperature were found among the three depths (F = 0.56, p > 0.05). However, the differences of temperature became obvious in the late growing seasons. The soil moisture significantly increased with soil depth (F = 4.59, p < 0.05), and the two-year mean values were 0.24, 0.25 and  $0.26 \text{ cm}^3 \text{ cm}^{-3}$  in the layers of 0–10, 10–20 and 20–40 cm, respectively (Fig. 1b). Irrigation temporarily increased soil moisture by 38-83%, 28-63% and 13-57% (relative terms) in the 0-10, 10-20 and 20-40 cm layers, respectively. The fluctuation in the gas diffusion coefficient decreased with greater soil depth and was influenced by irrigation or rainfall events (Fig. 1c). The gas diffusion coefficient significantly decreased with soil depth (F = 3.68, p < 0.05), and the twoyear mean values of soil gas diffusion were  $4.00 \times 10^{-7}$ ,  $2.52 \times 10^{-7}$ and  $1.67 \times 10^{-7} \text{ m}^2 \text{ s}^{-1}$  in the 0–10, 10–20 and 20–40 cm layers, respectively.

In the 0–10 cm layer, the application of urea significantly increased the soil mineral N contents (Table 1). Soil NO<sub>3</sub><sup>-</sup> contents significantly increased with increasing N fertilization rates, and the two-year mean values were 3.93, 5.63 and 8.15 mg N kg<sup>-1</sup> soil for the 0, 160 and 320 kg N ha<sup>-1</sup> treatments, respectively (Fig. 2a). The soil NH<sub>4</sub><sup>+</sup> content was significantly higher in the 320 kg N ha<sup>-1</sup> treatment than in the 0 and 160 kg N ha<sup>-1</sup> treatments (p < 0.05), and there was no significant difference in the NH<sub>4</sub><sup>+</sup> content between the 0 and 160 kg N ha<sup>-1</sup> treatments (p > 0.05) (Fig. 2b).

The root biomass was significantly higher in the 160 and  $320 \text{ kg N ha}^{-1}$  treatments than in the  $0 \text{ kg N ha}^{-1}$  treatment (p < 0.05), but there was no significant difference in root biomass between the 160 and 320 kg N ha<sup>-1</sup> treatments (p > 0.05) (Table 2). The cotton biomass significantly increased with the application of N fertilizer (p < 0.05), and the two-year mean values for the rates of 0, 160 and 320 kg N ha<sup>-1</sup> were 578.8, 851.7 and 1142.6 g C m<sup>-2</sup>, respectively (Table 2).

## 3.2. Soil-surface $CO_2$ efflux and $CO_2$ production

During the observation period, the temporal variations in the soilsurface CO<sub>2</sub> efflux (Fig. 3), CO<sub>2</sub> concentrations at different depths (Fig. 4) and CO<sub>2</sub> production in the soil profile (Fig. 5) were similar among the different treatments and years, increasing from May to August and then gradually decreasing according to the variations in soil temperature. Irrigation temporarily decreased the soil-surface CO<sub>2</sub> effluxes by 19–63%

Table 1

Results of repeated measures ANOVAs of the effects of N fertilization rates on the soil mineral N content in the 0–10 cm layer, soil-surface  $CO_2$  efflux and soil  $CO_2$  production in different soil layers in 2014, 2015 and both years combined.

Variables	Fertiliz	Fertilization treatments						
	2014		2015		Both			
	F	Р	F	Р	F	Р		
Soil NO <sub>3</sub> <sup>-</sup> content Soil NH <sub>4</sub> <sup>+</sup> content Soil-surface CO <sub>2</sub> efflux Soil CO <sub>2</sub> production (0-10  cm) Soil CO <sub>2</sub> production (10-20  cm)	11.40 2.22 1.08 1.18 1.93	< 0.01 > 0.10 > 0.10 > 0.10 > 0.10	8.66 1.66 1.76 1.62 1.43	< 0.01 > 0.10 > 0.10 > 0.10 > 0.10	20.22 3.86 2.73 2.74 2.98	< 0.01 < 0.05 0.07 0.07 0.06		
Soil CO <sub>2</sub> production (20–40 cm)	0.47	> 0.10	0.16	> 0.10	0.00	> 0.10		



**Fig. 2.** Temporal variations in soil nitrate (NO<sub>3</sub><sup>-</sup>) and ammonium (NH<sub>4</sub><sup>+</sup>) content in the 0–10 cm layer under three fertilizer rates in 2014 and 2015. *Error bars* for the soil NO<sub>3</sub><sup>-</sup> or NH<sub>4</sub><sup>+</sup> content represent the standard errors of the means (n = 3).

(Fig. 3), but its effect on the CO<sub>2</sub> production rates varied with the soil layers (Fig. 5). Irrigation did not affect the CO<sub>2</sub> production rate in the 0–10 cm layer but inhibited CO<sub>2</sub> production rates by 87–117% and 47–108% in the 10–20 and 20–40 cm layers, respectively. The production of CO<sub>2</sub> significantly decreased with soil layers (F = 152.17, p < 0.01), and the two-year mean values for the N treatments were 1.06, 0.38 and 0.21 g C m<sup>-2</sup> day<sup>-1</sup> in the 0–10, 10–20 and 20–40 cm layers, respectively. Overall, during the observation period, the production of CO<sub>2</sub> for the all treatments mainly occurred in the 0–10 cm layer, which accounted for 67% and 63% of the total production in the soil profile (0–40 cm) in 2014 and 2015, respectively (Table 3).

Compared with the simple functions, multiple regressions combining soil temperature and moisture improved the fit of the model, resulting in the lowest AIC values, relative high R<sup>2</sup> values and low RMSE values (Table 4). The multiple regression analysis showed that a higher soil-surface CO<sub>2</sub> efflux occurred at high soil temperatures and a certain range of soil moisture  $(0.21-0.23 \text{ cm}^3 \text{ cm}^{-3})$  (Fig. 6a), but the interactive effects of soil temperature and moisture on CO<sub>2</sub> production varied among the soil layers. In the 0-40 cm layer, CO<sub>2</sub> production exponentially increased with increasing temperature (Fig. 6b-d), although the temperature effect on soil CO<sub>2</sub> production decreased with increasing soil depth (Table 4). The effect of soil moisture on soil CO<sub>2</sub> production varied among the soil layers (Fig. 6b-d). Soil CO<sub>2</sub> production was positively correlated with soil moisture in the 0-10 cm layer but negatively correlated with soil moisture in the 10–20 cm layer. In the 20-40 cm layer, no significant correlation was found between soil  $CO_2$  production and moisture (p > 0.05). Overall, we observed higher rates of CO<sub>2</sub> production under conditions of both high soil temperature

and moisture in the 0–10 cm layer, but in the 10–20 cm layer, higher  $CO_2$  production occurred under high soil temperature and low soil moisture conditions (Fig. 6b and c).

## 3.3. Effect of N fertilization on soil-surface CO<sub>2</sub> efflux and CO<sub>2</sub> production

When the two years (2014 and 2015) were analysed together, N fertilization significantly increased the daily soil-surface CO<sub>2</sub> efflux at alpha = 10% (Table 1). We found a significantly positive correlation between soil-surface CO<sub>2</sub> efflux and soil NO<sub>3</sub><sup>-</sup> content in 2015 (Fig. 7c), but no significant correlations were found between soil-surface CO<sub>2</sub> efflux and the soil NH<sub>4</sub><sup>+</sup> contents in all treatments (Fig. 7b and d). The cumulative soil-surface CO<sub>2</sub> effluxes significantly (p < 0.05) increased with increasing N fertilization rates (Table 3), and the two-year mean cumulative effluxes obtained with the 0, 160 and 320 kg N ha<sup>-1</sup> treatments were 286.8, 315.4 and 360.9 g C m<sup>-2</sup>, respectively.

Urea application significantly enhanced the daily rates of CO<sub>2</sub> production in the 0–20 cm layer of the soil profile at alpha = 10%, but did not affect CO<sub>2</sub> production rates in the 20–40 cm layer (Table 1). Although N fertilization significantly (p < 0.05) increased the cumulative CO<sub>2</sub> production in the 0–40 cm layer, the effect of fertilization on CO<sub>2</sub> production varied among the soil layers (Table 3). In the 0–10 cm layer, the application of urea significantly (p < 0.01) accelerated the production of CO<sub>2</sub>, and the two-year mean values obtained for the 0, 160 and 320 kg N ha<sup>-1</sup> treatments were 151.07, 165.49 and 180.46 g C m<sup>-2</sup>, respectively. In the 10–20 cm layer, the olkg N ha<sup>-1</sup>

Table 2

Root and total biomass of cotton (g C m<sup>-2</sup>) under three fertilization rates in 2014 and 2015. The different letters indicate statistically significant differences between the three treatments (LSD test, p < 0.05). The values are expressed as the means  $\pm$  standard errors of the means (n = 3).

N fertilizer rate	Root biomass		Total biomass		
	2014	2015	2014	2015	
$0 \text{ kg N ha}^{-1}$ 160 kg N ha <sup>-1</sup> 320 kg N ha <sup>-1</sup>	73.0 ± 6.5 a 110.6 ± 10.3 b 114.9 ± 14.8 b	86.4 ± 6.5 a 105.4 ± 5.1 b 112.5 ± 13.4 b	544.6 ± 17.0 a 893.0 ± 39.5 b 1180.9 ± 109.2 c	612.9 ± 54.7 a 810.4 ± 39.5 b 1104.2 ± 58.2 c	



Fig. 3. Temporal variations in soil-surface  $CO_2$  effluxes under different fertilizer rates in 2014 and 2015. *Error bars* for the soil  $CO_2$  effluxes represent the standard errors of the means (n = 3).

treatment compared with the 160 and 320 kg N ha<sup>-1</sup> treatments, and a significant difference in CO<sub>2</sub> production was found between the 160 and 320 kg N ha<sup>-1</sup> treatments in 2015. In the 20–40 cm layer, there were no significant differences in cumulative CO<sub>2</sub> production among the three fertilizer treatments.

#### 4. Discussion

Nitrogen fertilization increased the soil-surface  $CO_2$  effluxes during the observation periods, and this result is consistent with previous findings (Bhattacharyya et al., 2012; Fuß et al., 2011; Luo et al., 2016;



Fig. 4. Temporal variations in soil  $CO_2$  concentrations at 0, 10, 20 and 40 cm depths under different fertilizer rates in 2014 and 2015. *Error bars* for the soil  $CO_2$  concentrations represent the standard errors of the means (n = 3).



Fig. 5. Temporal variations in  $CO_2$  production in the 0–10, 10–20 and 20–40 cm soil layers under different fertilizer rates in 2014 and 2015. *Error bars* for the soil  $CO_2$  production represent the standard errors of the means (n = 3).

#### Table 3

Cumulative soil-surface $CO_2$ efflux and $CO_2$ production (g C m <sup>-2</sup> ) in the soil profile under three fertilization rates in 2	2014 and 2015. The different letters denote significant differences
among the three treatments (LSD test, $p < 0.05$ ). The values are expressed as the means $\pm$ standard errors of the n	means $(n = 3)$ .

N fertilizer rate	Cumulative soil-surface CO <sub>2</sub> efflux	Cumulative CO <sub>2</sub> production					
		Soil layer					
		0–10 cm	10–20 cm	20–40 cm	0–40 cm		
2014 0 kg N ha <sup>-1</sup> 160 kg N ha <sup>-1</sup> 320 kg N ha <sup>-1</sup>	269.9 ± 9.0 a 300.8 ± 11.1 b 349.3 ± 4.3 c	150.9 ± 12.7 a 165.8 ± 15.7 ab 178.0 ± 6.0 b	40.2 ± 10.0 a 57.8 ± 1.8 ab 68.1 ± 13.2 b	30.9 ± 5.1 a 30.1 ± 6.4 a 22.6 ± 9.0 a	$\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$		
2015 0 kg N ha <sup>-1</sup> 160 kg N ha <sup>-1</sup> 320 kg N ha <sup>-1</sup>	303.6 ± 2.7 a 330.1 ± 4.4 b 372.5 ± 2.9 c	151.3 ± 3.8 a 165.2 ± 6.3 b 182.9 ± 4.0 c	$54.0 \pm 5.2 a$ $69.0 \pm 5.0 b$ $87.1 \pm 2.0 c$	25.5 ± 6.9 a 27.7 ± 1.2 a 30.4 ± 7.7 a	230.8 ± 3.2 a 261.9 ± 2.9 b 300.4 ± 11.0 c		

\* and \*\*: significant at p < 0.05 and p < 0.01, respectively.

Lv et al., 2014; Shao et al., 2014). The application of urea increases plant production and enhances the translocation of photosynthate to the rhizosphere, which increases root respiration (Guo et al., 2015; Hamada and Tanaka, 2001; Oh et al., 2005). However, Ding et al. (2010) found that N addition reduced root respiration in a maize field through decreased root biomass, because fertilization reduced the belowground allocation of photosynthate. In this study, the root biomass of cotton increased following urea application, which probably resulted in higher root respiration (Scheer et al., 2013; Shao et al., 2014). Additionally, N fertilization eliminates soil mineral N limitation, thereby indirectly increasing soil respiration (Ding et al., 2010; Kuzyakov et al., 2000; Luo et al., 2016). However, the soil  $NO_3^-$  content but not the soil  $NH_4^+$  content has been shown to affect soil respiration because it is the microbial community associated with nitrification, but not associated with ammonification, that regulates heterotrophic respiration (Luo et al., 2016). In this study, our results were partly consistent with those reported by (Luo et al., 2016), and we found a positive correlation between soil respiration and soil  $NO_3^-$  content in 2015. Overall, both root biomass and soil  $NO_3^-$  content increased following urea application in this study, probably resulting in enhanced root (Scheer et al., 2013; Shao et al., 2014) and heterotrophic (Luo et al., 2016) respiration rates. However, we could not distinguish the effects of N fertilization on

#### Table 4

Relationships between soil CO<sub>2</sub> efflux or production and soil temperature and moisture using simple and multiple functions. *SE* is the soil-surface CO<sub>2</sub> efflux, and *SP*<sub>0-10</sub>, *SP*<sub>10-20</sub> and *SP*<sub>20-40</sub> are the amounts of CO<sub>2</sub> produced in the 0–10, 10–20 and 20–40 cm layers, respectively (g C m<sup>-2</sup> day<sup>-1</sup>). The  $Q_{10}$  hyperbolic and linear functions are  $R = R_{10} \cdot Q_{10}^{(T-10)/10}$ ,  $R = a + b \cdot M + c / M$  and  $R = a + b \cdot M$ , respectively. *R* is CO<sub>2</sub> efflux or production (g C m<sup>-2</sup> day<sup>-1</sup>);  $R_{10}$  is CO<sub>2</sub> efflux or production at 10 °C (g C m<sup>-2</sup> day<sup>-1</sup>);  $Q_{10}$  is the temperature sensitivity of the soil CO<sub>2</sub> efflux or production; *T* and *M* are the soil temperature (°C) and moisture (cm<sup>3</sup> cm<sup>-3</sup>), respectively, of different layers; and *a*, *b* and *c* are functional coefficients.

Variable	Function	R <sub>10</sub>	Q <sub>10</sub>	а	b	c	R <sup>2</sup>	RMSE (%)	AIC	Р
Soil temperature										
SE	Q <sub>10</sub>	0.54	3.23				0.66	15.6	- 109.8	< 0.01
SP <sub>0-10</sub>	Q <sub>10</sub>	0.26	3.19				0.55	11.2	-240.2	< 0.01
SP <sub>10-20</sub>	Q <sub>10</sub>	0.06	5.68				0.25	24.7	-235.1	< 0.01
$SP_{20-40}$	Q <sub>10</sub>	0.03	6.91				0.12	30.1	- 286.7	< 0.01
Soil moisture										
SE	Hyperbolic			- 4.01	10.16	0.83	0.03	48.1	-1.2	< 0.05
SP <sub>0-10</sub>	Linear			-0.25	5.43		0.18	25.2	-186.1	< 0.01
SP <sub>10-20</sub>	Linear			1.61	- 4.91		0.15	39.7	-270.0	< 0.01
SP <sub>20-40</sub>	Linear			0.36	-0.61		0.00	44.1	- 343.0	> 0.05
Soil temperature and moisture										
SE	Q <sub>10</sub> and hyperbolic	0.55	3.17	4.52	-8.38	-0.36	0.64	18.0	- 141.9	< 0.01
SP <sub>0-10</sub>	Q <sub>10</sub> and linear	0.26	3.36	0.22	3.07		0.61	12.1	- 291.2	< 0.01
SP <sub>10-20</sub>	Q <sub>10</sub> and linear	0.13	4.14	2.17	- 6.56		0.30	32.6	- 296.7	< 0.01
SP <sub>20-40</sub>	$Q_{10}$ and linear	0.11	4.40	0.69	- 1.45		0.07	41.3	- 350.5	< 0.01

a 0 cm









d 20-40 cm



Fig. 6. Relationships of soil-surface  $CO_2$  efflux (a) or production in 0–10 (b), 10–20 (c) and 20–40 (d) cm soil layers with soil temperature and moisture. The black and grey dots denote the soil-surface  $CO_2$  efflux or production higher or lower than the values simulated the multiple regression models.



**Fig. 7.** Relationships between the soil-surface  $CO_2$  efflux and the soil mineral N ( $NO_3^-$  and  $NH_4^+$ ) content in the 0–10 cm layer under different fertilization rates (0, 160 and 320 kg N ha<sup>-1</sup>) in 2014 (a and c) and 2015 (b and d).

root and soil microbial respiration because there likely were interactive effects of these factors on root and heterotrophic respiration. For example, Moinet et al. (2016) suggested that the response of heterotrophic respiration to N addition is mediated by the presence of roots because a reduction in root biomass enhanced the competitiveness of soil microorganisms, thereby increasing heterotrophic respiration. Therefore, further studies should investigate the effects of N fertilizer application on root and heterotrophic respiration separately using the  $\delta^{13}$ C technique (Moinet et al., 2016).

In general, soil respiration increases with increasing soil temperature below 35 °C (Lloyd and Taylor, 1994), as long as soil moisture is not a limiting factor (in the range from 0.20 to  $0.30 \text{ cm}^3 \text{ cm}^{-3}$ ) (Gaumont-Guay et al., 2006). Many researchers have described the relationships between soil temperature and surface-soil CO2 efflux and CO<sub>2</sub> production rates using the Q<sub>10</sub> model (Gaumont-Guay et al., 2006; Hashimoto and Komatsu, 2006; Koncz et al., 2015a), and our results showed that soil temperature is significantly correlated with soil-surface CO<sub>2</sub> efflux and CO<sub>2</sub> production rates. However, the effect of soil temperature on soil CO<sub>2</sub> production varied depending on the soil layer, and this finding likely resulted from the regulation of the vertical distribution of CO<sub>2</sub> production rates by soil biological activities, which were highly influenced by the soil temperature. In the soil profile (0-40 cm layer), a large portion (63-67%) of the soil-surface CO<sub>2</sub> efflux was produced in the 0-10 cm layer. Similar results were obtained in previous studies (Guo et al., 2015; Nan et al., 2016), which attributed greater soil respiration rates to the higher plant root biomass and concentrations of organic matter and O<sub>2</sub> in the topsoil (Guo et al., 2015; Hamada and Tanaka, 2001; Oh et al., 2005). In our study, SOC decreased with increasing depth, as mentioned in Section 2.1, which implies higher CO<sub>2</sub> production in the topsoil due to microbial decomposition. Although we did not measure the vertical distribution of the root biomass, Zhao et al. (2010) found that the root length density in this cotton field decreases with increasing soil depth and that a large portion of the roots are distributed in the 0-10 cm layer.

Many researchers have reported that soil-surface  $CO_2$  effluxes, which reflect soil biological activities, increase with increasing soil water content, but excessive soil moisture depresses soil respiration by limiting the transport of  $CO_2$  or  $O_2$  in the soil profile (Gaumont-Guay et al., 2006; Yan et al., 2013). In this study, higher soil water contents following irrigation events temporarily restricted soil-surface  $CO_2$  effluxes and  $CO_2$  transport in the soil profile, which is consistent with the results of previous studies in forest and agricultural soils (Hashimoto and Komatsu, 2006; Nan et al., 2016; Pumpanen et al., 2008). The higher  $CO_2$  concentration can mainly be attributed to water-blocked soil pores and reduced diffusivity, which result in the accumulation rather than transport of  $CO_2$  in the soil profile (Gaumont-Guay et al., 2006; Pumpanen et al., 2008).

We found that  $CO_2$  production in the 0–10 cm layer of the soil profile was not restricted by excessive soil water content, and a similar result was reported by Hashimoto and Komatsu (2006), who found that a sudden increase in soil moisture in the 0–10 cm layer did not change the rates of  $CO_2$  production. However, in the 10–20 cm layer, we found that  $CO_2$  production was markedly reduced under high soil moisture conditions. These differences are probably attributable to the following: (1) the  $O_2$  diffusion rate in the topsoil might remain relatively high under high soil moisture conditions, whereas the  $O_2$  diffusion rate in the deeper soil may be considerably lower and act as a limiting factor to soil respiration; (2) although Hashimoto and Komatsu (2006) showed that an insufficient amount of dissolved  $CO_2$  in the water of boreal forest soils, there was a significant amount in saline/alkaline soil water (Li et al., 2015). Therefore, in the 10–20 cm layer, where the  $CO_2$ production rate was relatively low, the change in the calculated  $CO_2$  production rate was likely dominated by abiotic processes (e.g., degassed or dissolved  $CO_2$ ) rather than actual soil respiration.

It is worth noting three disadvantages related to the use of the gradient method applied in this study. First, although soil porosity decreases with time after tillage, it is usually treated as a constant value for a given field, but Han et al. (2014) found that soil porosity decreased within 50 days after tillage and then remained relatively stable. In this study, we likely underestimated the calculated soil CO2 production in the early observation period (May) by assuming a constant soil porosity. Second, this gradient method assumes that diffusion is the main mechanism of  $CO_2$  transport in the soil (Rey, 2015) and does not consider convective movement or the transport of CO<sub>2</sub> dissolved in soil water. However, Pumpanen et al. (2008) suggested that no significant CO<sub>2</sub> transport occurs in the soil water because the movement of soil water is very slow. Finally, previous studies have used continuous halfhourly measurements of CO<sub>2</sub> concentrations to calculate the CO<sub>2</sub> production in the soil profile (Guo et al., 2015; Han et al., 2014; Jassal et al., 2005; Pumpanen et al., 2008); thus, field data should be collected more frequently because the weekly observations in this study could not discern continuous changes, which likely led to a misestimation of the cumulative CO2 production. Despite these limitations, understanding the CO<sub>2</sub> transport and production in the soil profile is important for studying the mechanisms of soil respiration and improving our understanding of the interactions between management practices and soil respiration.

#### 5. Conclusions

In the oasis cotton fields of our study, greater CO<sub>2</sub> production was found to occur in the 0-10 cm soil layer, and this production amount accounted for 63% to 67% of the total amount of CO<sub>2</sub> generated in the soil profile (0-40 cm). The application of N fertilizer increased the soilsurface CO<sub>2</sub> efflux by enhancing the CO<sub>2</sub> production rates in the 0-20 cm layer. We observed higher soil-surface CO<sub>2</sub> efflux under high soil temperature and a certain range of soil moisture conditions  $(0.21-0.23 \text{ cm}^3 \text{ cm}^{-3})$ . In the 0-40 cm layer, CO<sub>2</sub> production exponentially increased with increasing temperature, although the temperature effect on soil CO<sub>2</sub> production decreased with increasing soil depth. Soil CO<sub>2</sub> production was positively correlated with soil moisture in the 0-10 cm layer but negatively correlated with soil moisture in the 10-20 cm layer. Overall, our results suggest that the impact of management practices and soil environmental variables on CO<sub>2</sub> production varied depending on the soil layer. Further studies should separate the "real"  $CO_2$  production rate and the abiotic  $CO_2$  processes in the saline/ alkaline soil profile (e.g., the dissolved or degassed CO<sub>2</sub>).

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